BIOMONITORING NETWORK REPORT OF WAHDIENGLIENG STREAM



ST. ANTHONY'S COLLEGE SHILLONG, MEGHALAYA, INDIA 2014

The Biomonitoring Network is an outcome of the training program on "Capacity Building for Freshwater Ecosystem Conservation in Northeastern India" organized by Shillong College, Shillong on $14^{th} - 21^{st}$ July 2014, in collaboration with **Rufford Foundation for** Nature Conservation (UK), Global Water for Sustainability Program, Florida International University (USA) and Foundation for Ecological Research, Advocacy and Learning (FERAL), Pondicherry, India. The participants of the training program included students and teachers from different colleges of Shillong, Meghalaya. During the training, the participants were taught how to assess and monitor the quality of water in the stream or river based on the presence of macroinvertebrates and also on the physico-chemical parameters. At the end of the training, it was decided that each college will adopt one stream in and around Shillong city and monitor the water quality of that particular water body. Dr. Sonali Saha who is the Co-Principal Investigator and Coordinator of Science Programs, Touro College, South, Miami Beach, Florida, USA, and who was the main instructor during the training program is heading the Network. She wrote a letter to all the Principals of the colleges in Shillong, requesting them to be part of the Biomonitoring Network. As a result of the initiative of Dr. Sonali Saha and also of Dr. Bashida Massar of the Department of Zoology St. Anthony's college who attended the training program, a team is set up in the college in August 2014 with the following as members:

Name	Designation	Department
Dr. Bashida Massar	Assistant Professor	Zoology
Mr. Rupak Nath	Assistant Professor	Fishery Science
Ms. Ebelmon Nongbri	Assistant Professor	Botany
Ms. Dawanlangki Shadap	B.Sc. Third year Student	Zoology
Ms. Sumera Sangma	B.Sc. third year Student	Zoology
Ms. Ibalarisha Donshiew	B.Sc. second year student	Fishery Science
Mr. Mayborn Dkhar	B.Sc. second year student	Fishery Science
Mr. Friendly Majhong	B.Sc. second year student	Fishery Science

St. Anthony's College Biomonitoring Network Team

St. Anthony's College Biomonitoring Network Team adopts Wahdienglieng, a small stream in Risa Colony, Shillong near Zoological Survey of India (ZSI).

Biomonitoring of Wahdienglieng stream, Shillong, Meghalaya, India

Rationale: Aquatic insects vary in their sensitivity to changes in water flow and quality; the more sensitive organisms are thus good indicators of water quality and watershed ecosystem health. Flow alterations and soil erosion can lead to the disappearance of many insect groups. By knowing what insect communities exist in pristine streams in Meghalaya, one can infer the condition of other streams in the region.

Aim: To monitor the quality of a stream or river

Objectives: This project has to meet the following objectives:

- Introduce students to the concept of biological monitoring of water quality, based on the presence, abundance and diversity of aquatic macroinvertebrates (insect larvae and other taxa).
- To create a field guide of aquatic invertebrates in Meghalaya over a year (July 2014-June 2015), for identification at the order level and further levels (family, genus if possible).
- To be part of the biomonitoring network of faculty and students from various institutions in the region.

Report on field survey (22.08.2014)

Study Site

We selected Wahdienglieng stream as the study site, and conducted surveys of macroinvertebrate diversity and abundances in conjunction with water quality monitoring during the monsoon and early winter season. Wahdienglieng is about 2 km from St. Anthony's college, Shillong (Fig. 1; N - 25^{0} 33.723', E - 91^{0} 53. 606'), located at the elevation of 1515 m to 1520 m asl.

The immediate catchment of the stream is forested; however being in an urban area, has considerable human use and impacts both directly in-stream (activities such as washing clothes and utensils) as well as across the catchment (sedimentation, pollution in storm water runoff). On this day, few studies were performed including pH, alkalinity and the organisms inhabiting the stream. Some people were found to wash their clothes in this stream and some boys were also fishing and caught *Channa stuarti* a particular species of *Channa* which inhabits only in high altitude.

Based on the surveys the impact of human activities was evident. The clarity of water was medium to poor, and few species of aquatic macroinvertebrates were found. The species such as blackfly larvae are indicators of poor water quality.



Figure 1: Study site in a stream within a forested riparian area surrounded by a mosaic of urban settlements.

Water quality analysis (methods)

Water quality is the term used to describe the chemical, physical and biological characteristics of water. Fish as well as all aquatic organisms can survive, adjust and adapt to

some fluctuations in the water quality, but they suffer and die once the limits have been crossed. Therefore, it is important to assess the prevailing quality of the water in the study area.

Collection of water samples

Water samples were collected from the lake (pollution-exposed) and control water body in sterilized plastic containers having a capacity of 5 litres and brought to the laboratory for analysis of different parameters. Both physical and chemical parameters were analyzed. The method of water collection was done as per the Standard methods for the examination of water and wastewater, American Public Health Association (APHA), 1992.

Hydrogen Ion Concentration (pH)

pH is a measure of the acidity and alkalinity of the water. It is measured by using pH universal indicator solution (pH 4 - 10) and pH meter. Water was taken in a test tube and few drops of the pH indicator were added. The colour produced is compared with the standard colour and the pH was recorded. Readymade buffers were used for setting the pH meter and then the measurement of samples were calibrated out and recorded (Verma and Srivastava, 1996).

Dissolved Oxygen (DO)

Dissolved oxygen is the amount of gaseous oxygen dissolved in an aqueous solution and is expressed as a concentration in terms of milligrams per litre (mg/L). Dissolved oxygen is determined by Winkler's Iodometric method (Verma and Srivastava, 1996). Water samples were collected in 250 ml glass Biological Oxygen Demand (BOD) stoppered bottle up to the brim. During collection of samples, precautions were taken not to allow any kind of air bubbles to enter inside the bottle. 1 ml each of MnSO₄ and alkaline KI solutions were added to the sample very carefully by the side of the bottle. Two separate pipettes were used for the reagents. The bottle was closed with the stopper and mixed by inverting several times. A brownish-orange cloud of precipitate appeared. When this precipitate has settled to the bottom, the sample was mixed by turning it upside down several times and let it settle again. 1 ml of concentrated sulfuric acid via a pipette held just above the surface of the sample was added and shaken well to dissolve the precipitate. 25 ml of the content of the bottle was carefully transferred to the conical flask for titration. The content was titrated against sodium thiosulphate solution using starch as an indicator. At the end point, initial dark blue colour became colourless.

Calculation:

 $DO (mg/L) = \frac{8 \text{ x Volume of Titrant used x 1000 x Strength of Titrant}}{Volume of sample taken}$

Free Carbon Dioxide

Free carbon dioxide was determined following the APHA standard method (1992). 50 ml of the water sample was taken in a beaker and 2 - 3 drops of phenolphthalein indicator was added to it. Since no pink colour developed, the solution was titrated against 0.045 N Sodium carbonate solution till a faint permanent pink colour appears and the reading was recorded.

Calculation:

Free CO₂ (mg/L) = $\frac{\text{Volume of Titrant used x 1000}}{\text{Volume of sample taken}}$

Alkalinity

Alkalinity of the water was determined following APHA standard method (1992). 50 ml of the water sample was taken in the conical flask and to it 2-3 drops of phenolphthalein indicator was added. 2 - 3 drops of methyl orange indicator was then added and the solution turned yellowish in colour. It was then titrated with 0.02 N H_2SO_4 till the yellow colour turned orange and the reading was recorded.

Calculations:

Alkalinity (mg/L) = $\frac{\text{Volume of Titrant used x 1000}}{\text{Volume of sample taken}}$

STUDY FINDINGS

Temperature

Seasonal temperature fluctuations are evident in the measurements, with water temperature varying from 19 C in the monsoon season to 10 C in winter.

Width of stream

Stream dimensions did not vary across the seasons. On average the stream was 3 m wide and 0.5 m deep, 0.4 - 0.6 m.

Size of rock boulders: 0.4 - 0.7 m length or diameter

Water Turbidity (Transparency of water in column using Secchi Disk):

Point of appearing (D1); Point of disappearing (D2) Spot 1: D1 = 13 cm; D2 = 13.1 cm Spot 2: D1 = 10 cm; D2 = 13.5 cm Spot 3: D1 = 7 cm; D2 = 13.2 cm Transparency = (D1 + D2) / 2, Turbidity when expressed as NTU was 90, suggesting that water is quite murky.

Stream Velocity (using Float method):

Stream velocity was 40 cm s⁻¹ in the monsoons, and was not measured in the winter.

Macroinvertebrates:

We mainly found Blackfly larvae (many of them under the stones). Flatworms commonly called planarians are also found under some stones. In addition there were caddisfly, mayfly larvae and water penny beetle under some stones.

Hydrogen ion concentration (pH)

The pH values were comparable across the seasons and were found to be steady around 7 to 7.5 (Table 1).

Dissolved Oxygen (DO)

Dissolved oxygen was correlated with temperature, and greater concentration of DO was recorded in winter (14-16 mg L^{-1}) compared to monsoons (6.4 – 8 mg L^{-1})(Table 1).

Free carbon dioxide

Concentration of freeCO₂ was greater in winter than the monsoons 8 mg L^{-1} in monsoons and 10 mg L^{-1} in winter (Table 1).

Total Alkalinity

Alkalinity measures the total amount of base present and indicates a pond's ability to resist large pH changes, or the buffering capacity. The most important components of alkalinity are carbonates and bicarbonates. Alkalinity was greater in monsoons 18 mg L^{-1}) compared to the winter (16 mg L^{-1} ; Table 1).

DISCUSSION AND CONCLUSIONS

Wahdienglieng stream has a swift flow on account of being a hill stream (flowing down the hill slope), and this is reflected in moderately high dissolved oxygen (DO). The DO in the stream was moderate in the monsoon season and higher in winter, as the DO is a factor of temperature as well as flow. However, despite the moderately high DO, the stream was found to have low macroinvertebrate richness and diversity. The invertebrate community was dominated by blackfly larvae which thrive in water with moderate to fast currents as they are the filter feeders of organic substances suspended in water. The stream appears to be impacted by disturbance (human activities such as washing utensils and clothes, and possibly grey water discharge) as few niches of the macroinvertebrate food web seem to be occupied. Possible causes of the observed low invertebrate diversity can be sediment that covers up streambed rocks thereby removing habitat for invertebrates, high nutrients in laundry detergent, kitchen waste and wastewater that can lead to periodic algal blooms. To understand this low diversity, further observations are necessary.

Parameters	Monsoon	Winter
рН	7 - 7.5	7.5
Dissolved Oxygen (mg L ⁻¹)	6.4 - 8	14 - 16
Free $CO_2(mg L^{-1})$	8	10
Total Alkalinity (mg L ⁻¹)	18 - 18.4	14 - 16
Temperature (⁰ C)	18 - 20	10
Velocity (cm s ⁻¹)	40	-
Transparency (NTU)	90	-
Channel width (m)	3.07 - 3.16	-
Channel depth (m)	0.49 - 0.58	-

 Table-1.Physico-chemical characteristics of Wahdienglieng recorded in the months of

 September- October (monsoon season) and November – December (winter season)



Figure 2:Wahdienglieng stream in Risa Colony, Shillong





Figure 3: Activities going on in the stream by local people

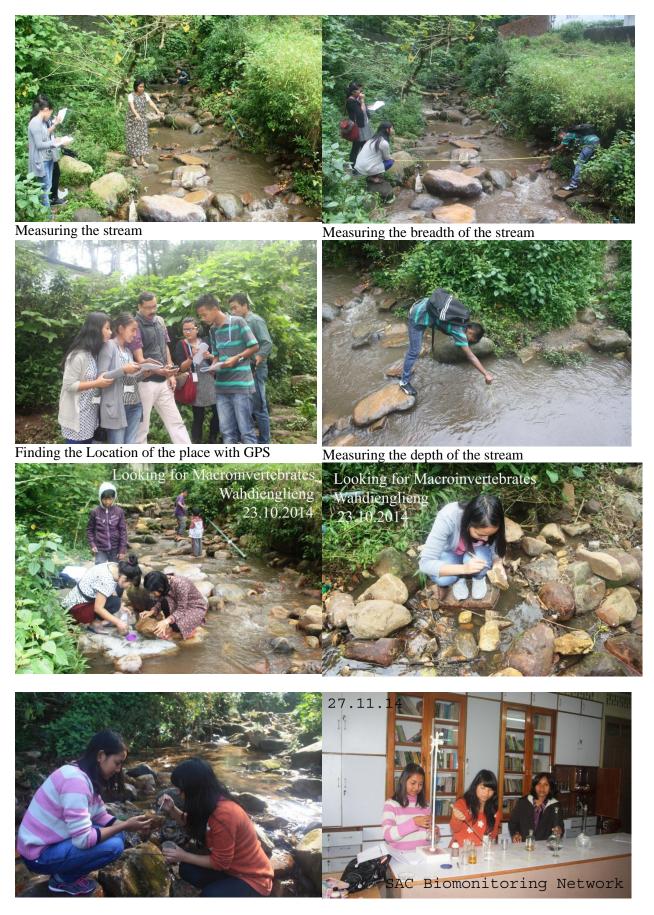


Figure 4: Activities performed by the Biomonitoring Network team



Figure 5: Fauna of Wahdienglieng stream, Shillong

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